

Antibodies to Signaling Molecules and Receptors in Alzheimer's Disease are Associated with Psychomotor Slowing, Depression, and Poor Visuospatial Function

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Abstract.

Background: Alzheimer's disease (AD) is associated with several antibodies as well as signaling molecules and receptors. These may be detrimental in the presence of a disrupted blood-brain barrier (BBB).

Objective: To investigate whether the levels of antibodies toward 33 signaling molecules involved in neurotransmitter, vascular, and immune functions were associated with AD and, within the AD group; cognitive function and mood.

Methods: Antibodies in sera from patients with mild AD [$(n=91)$ defined as a Mini-Mental State Examination ≥ 20 or a Clinical Dementia Rating Scale ≤ 1] and healthy controls ($n=102$) were measured with enzyme-linked immunosorbent assays. Levels in AD and controls were compared by Mann-Whitney test. In the AD group, associations between antibodies and psychometric test scores were analyzed by robust regression. The false discovery threshold was set to 0.05.

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Results: Antibodies to serotonin receptors [5-HT_{2A}R (effect size (r)=0.21, p =0.004), 5-HT_{2C}R (r =0.25, p =0.0005) and 5-HT₇R (r =0.21, p =0.003)], vascular endothelial growth factor receptor 1 [VEGFR1 (r =0.29, p <0.001)] and immune-receptors (Stablin-1 (r =0.23, p =0.001) and C5aR₁ (r =0.21, p =0.004) were higher in AD. Psychomotor speed was associated with D₁R-abs (β 0.49, p <0.001), depression with ETAR-abs (β 0.31, p <0.001), and visuospatial function with 5-HT_{1A}R-abs (β 0.27, p =0.004) despite similar antibody levels compared to controls.

Conclusions: Antibody levels to VEGFR1, serotonergic receptors, and receptors in the immune system were increased in AD. Antibodies at similar levels as in controls were associated cognitive dysfunction and depression in AD.

Keywords: C5aR, 5-HT_{2A}R, 5-HT_{2C}R, 5-HT₇R, MADRS, naturally occurring antibodies, Stablin-1, Trail Making A, VEGFR1, VOSP

INTRODUCTION

Alzheimer's disease (AD) is the most common form of dementia [1]. Neuropathological hallmarks of AD are accumulation of amyloid- β (A β) in plaques and tau proteins in tangles [2] causing extensive neuronal cell death [3], synapse loss [4], and microglial activation [5]. Naturally occurring antibodies (Nabs) of the IgG type are ubiquitous in human sera [6], including brain reactive antibodies [7]. Their physiological functions include clearance of apoptotic cells [8]. Under physiological conditions, the concentration of immunoglobulin G (IgG) inside the brain is extremely low and most Nabs do not cause disease. However, Nabs directed at proteins in the brain with pathogenic potential do exist at low frequencies in healthy populations [9]. Their pathogenic effect on the brain is likely dependent on damage to the blood-brain-barrier (BBB) [10]. Most AD patients have cerebral amyloid angiopathy and microvascular disease in the brain. Due to the BBB impairment seen in AD, IgG penetration to the brain might be increased [11].

AD has previously been associated with antibodies toward A β , tau, vascular-related molecules, lipid molecules, neurotransmitter receptors, glial markers, and cellular enzymes (reviewed in [12]). Antibodies directed to the angiotensin 2 type 1 receptor (AT1R-abs) [13], the α_1 - and β_2 -adrenoceptors, glutamate, serotonin, dopamine, and the N-methyl-d-aspartate glutamate receptors (NMDAR-abs) are found in AD sera [14–16]. Antibodies to receptors and signaling molecules may have pathogenic effects beyond the Fc-receptor mediated effects, by inducing signal-transduction mechanisms and as such they could act on receptors involved in neurotransmission under conditions with BBB impairment. In this explorative study, we aimed to comprehensively investigate antibodies to vascular, immune, serotonergic, dopaminergic, and muscarinic cholinergic

receptors and signaling molecules to establish a) if they are increased in AD compared to healthy controls and b) if they are associated with cognitive functions and mood in AD.

MATERIAL AND METHODS

Subjects

Patients with mild dementia were recruited to a longitudinal cohort-study between 2005 and 2007 from three participating centers in Rogaland and Hordaland counties in Norway, the Dementia study in Western Norway (DemVest) [17]. From this larger cohort of 250 patients, 91 patients with AD and available blood samples were included in this study. Clinical data, psychometric test results, and measurements in sera from baseline constituted the main objects of study. Mild dementia was defined as a Mini-Mental State Examination (MMSE) test score ≥ 20 or a Clinical Dementia Rating scale = 1. AD was diagnosed according to the criteria from the National Institute of Neurological and Communicative Disorders and Stroke-Alzheimer's Disease and Related Disorder Association (NINCDS-ADRDA) [18]. A trained research clinician with long experience in geriatric psychiatry, neurology, or geriatric medicine conducted a detailed medical history and standardized clinical examination. Structural MRI was conducted for diagnostic purposes [19]. Patients were recruited for brain donation. Among the 91 cases included in this study, 19 patients underwent autopsy and 16 had a neuropathological diagnosis of AD (84.2%). Exclusion criteria were acute delirium or confusion, terminal illness, recently diagnosed major somatic illness, and previous bipolar or psychotic disorder. Patients were examined with neuropsychological tests (see below). The Cumulative Index Illness Rating Scale (CIRS, reviewed

in: [20]) was registered for patients and controls. CIRS scores co-morbidity in 14 system domains by a severity score that can be added to give a total score (CIRS-total). ApoE genotyping [21] and routine blood tests were performed, including leukocyte count as a general marker of inflammation (see "Statistics" below). Further details of the clinical and biomarker assessment program are described elsewhere [17].

One hundred and three healthy community-dwelling elderly controls were recruited from senior centers in Bergen, Hordaland, during 2014. The control group was frequency matched by design on age and gender compared to the AD group (see Table 1, results). Good physical and mental health and a MMSE test score ≥ 28 were inclusion criteria. A trained internist conducted a clinical interview and examination. Ongoing psychosis, delusions, depression, major medical illness, previous bipolar- or psychotic disorders were exclusion criteria. Cardiovascular risk factors, cardiovascular disease, cerebrovascular disease [transient ischemic attack or stroke (CBVD)], other known illnesses, CIRS, medication use, body mass index (BMI), and blood pressure were registered for both groups.

Ethics

Ethical approval was granted from the Regional Ethics Committee (REK approval no.: 2010/633). Both patients and controls provided written consent to participate in the study after procedures had been explained in detail, and in case of patients with AD, also to a caregiver [13].

Antibody measurements

Sera from patients and controls were collected at baseline and stored at minus 80°C. Sample labels were blinded for the laboratory prior to analyses. Recombinant signaling molecules and receptors were expressed in Chinese hamster ovary (CHO) cells purified from membranes and used as antigens in subsequent enzyme-linked immunosorbent assays (ELISA) (CellTrend GmbH, Luckenwalde, Germany) [22]. A more detailed description of the ELISA procedure can be found in the Supplementary Material with coefficients of variation (Supplementary Table 2) and a list of abbreviations (Supplementary Table 4).

Mean storage time was 1 year for controls and 7 years for patients. Bias due to different storage times was investigated. First, by dividing the AD group into

three subgroups based on mean years of storage time and comparing antibody levels in each of these subgroups (by year) to controls, a change in associations indicative of a temporal effect would indicate bias. Second, significant correlations between the levels of antibodies and storage time within the AD group would indicate bias.

Antigen selection

We selected antigens from a broad range of receptor and signaling systems expressed in the brain, vessels, immune system, lung, heart, and kidney that participate in key signal transduction pathways. We measured antibodies to three signaling molecules: vascular endothelial growth factor alpha (VEGFA); platelet derived growth factor (PDGF); and nerve growth factor (NGF); and twenty-two antibodies to receptors of the adrenergic-, dopaminergic-, serotonergic-, and cholinergic systems. Antibodies to the vascular receptors endothelin receptor type A (ETAR), protease-activated receptor 1 (PAR1), VEGF, VEGFR1, and VEGFR2 and antibodies to the innate immune system receptors [Stabilin-1 (Stab1) and Stabilin-2 (Stab2) and the complement component 5a receptor type 1 (C5aR₁)] were also measured [23]. Finally, measurement of antibodies to nerve growth factor (NGF, linked to neuronal survival [24]) and the receptor for advanced glycation end-products (RAGE) that transport A β from the blood to the brain was performed [25].

Neuropsychological testing and mood

Four specific cognitive domains were tested: verbal memory [List A, Short Delay Cued Recall, California Verbal Learning test-II (CVLT-II) subtest (CVLT-SDCR)], visuospatial function [Visual Object and Space Perception Battery – Silhouettes subtest (VOSP)], psychomotor speed [Trail Making A Test (TMT-A)], and executive function (Stroop color-word test). TMT-A is a measure of psychomotor speed [26]. The Stroop Color-Word test use interference to test the ability to shift attention (Stroop effect: Stroop-E) and it is a measure of executive function. Depression was assessed using the Montgomery and Aasberg Depression Rating Scale (MADRS), a clinical interview consisting of 10 items scored between 0 and 6 that has shown good test performance in AD [27]. An overview of central tendencies and dispersions of the psychometric scores can be found in Table 1, results.

Power analyses

We could not identify prior studies to identify effect sizes *a priori*. We therefore used small-to-moderate effect size for power analyses. Alpha levels of 0.01 were used as a crude adjustment to accommodate power loss due to adjustment for multiple testing (see Statistics). A sample size of 200 in equal groups was necessary to detect a moderate difference ($R=0.24$) in antibody levels between the groups with a power of 0.8 [alpha=0.01, Mann-Whitney U test (MWUT)]. The required sample size to detect an R^2 -increase of 0.13 from 1 predictor in multiple linear regression (regression with antibodies and psychometric test scores) with a power of 0.8 was 82 (alpha=0.01, predictors=5). Power to detect interactions in multivariate regression (not calculated) has been estimated to be 1/3 of the overall power, making our study underpowered to detect such effects [28], but multivariate analyses of confounding do not lead to a similar power loss [29]. Power analyses were conducted with G*Power 3.0 [29].

Statistics

p-values less than 0.05 were considered statistically significant and adjustment for multiple comparisons were made with false discovery rate set to 5% (Benjamini-Hochberg). Univariate tests were analyzed using non-parametric statistics and ROC analyses (evaluated at specificity and sensitivity of 0.7) due to skewed distributions of antibody levels. A clustered heatmap was made from Spearman correlations between antibody levels. In multivariate analyses, antibody levels were transformed to normality by the Rankit method (logistic regression) and Box-Cox transformations (linear regression). Cognitive test scores were log and square root transformed to normality and optionally inversed so that “higher is worse” was true for all.

Confounding effects due to differences between the AD and control groups were evaluated by comparing estimates of antibody association with AD by logistic regression. Estimates from a simple model with age and gender (model 1) was compared to estimates a model with age, gender, known hypertension, current systolic blood pressure >140 mmHg, diabetes, current smoking, cardiac disease, and CBVD as potential confounders (model 2). A 10% change in estimate [(Crude Odds Ratio (OR) – Adjusted OR) / Crude OR] was considered minor confounding

and change to a non-significant *p*-value, major confounding. Finally, we compared antibody levels between participants with neuropathologically confirmed AD to those with a clinical diagnosis of AD.

Potential linear relationships between antibodies and psychometric test scores were analyzed by standardized robust regression by MM estimation [30] (package “mmregress”, STATA) due to the presence of influential outliers (criteria: Cooks distance >4(N – k - 1) and/or leverage >2p/N). Due to strong correlations between antibodies (illustrated in Fig. 3), the assumption of non-multicollinearity was broken. Each antibody was thus entered separately with covariates to identify the strongest associations. We addressed major confounding of the strongest associations from ApoE genotype, leukocytes, vascular risk factors or CBVD (10 cases missing for ApoE genotype and total cholesterol were deleted list-wise, as they were missing completely at random).

Analyses were performed using SPSS Statistics version 22.0 (SPSS Inc., Chicago, IL, USA), STATA 14 (StataCorp. 2015. *Stata Statistical Software: Release 14*. College Station, TX: StataCorp LP), and Orange Data Miner (Orange, in collaboration with Open Source Community, Bioinformatics lab., University of Ljubljana, Slovenia) [31].

RESULTS

Study participants

The AD and control groups were frequency matched by design on age and gender with a good match. They were also well matched on BMI, ever smoking, and CIRS, but there were significant differences between the groups in the prevalence of cardiovascular risk factors and disease (Table 1).

Differences in antibody levels between AD and controls

Eleven out of 33 antibodies measured were significantly different between the groups and six antibodies were discoveries, after multiple testing was adjusted for (Fig. 1). Antibody levels by groups can be found in Supplementary Table 1.

VEGFR1-abs was most associated with AD ($r=0.28$, $p=0.0002$) with 5-HT_{2A}R-abs ($r=0.21$,

Table 1
Characteristics of the study participants

Variables	Alzheimer's disease versus healthy controls		
	N=91	N=103	p-values
Age (M [SD])	74.9 [7.8]	73.5 [7.5]	0.195 ^a
Female	27%	26%	0.843 ^b
MMSE (M [SD])	23.8 [2.19]	29.3 [0.95]	<0.001 ^{**a}
Previously diagnosed hypertension	45%	32%	0.044 ^{*,b}
Current systolic BP \geq 140	76%	33%	<0.001 ^{**b}
Cardiac disease ^d	25%	11%	0.026 ^{*,b}
Diabetes	9%	3%	0.077 ^b
Cerebrovascular disease ^e	15%	5%	0.014 ^{*,b}
Body mass index (M [SD])	23.8 [4.1]	24.6 [3.5]	0.170 ^a
Present smoker, %	24%	11%	0.013 ^{*,b}
Ever smoker, %	40%	40%	0.460 ^b
CIRS total (Mdn [IQR])	3 [4]	3 [3.3]	0.381 ^c
Alzheimer's disease only			
MADRS, M [SD]		6.7 [5.6]	
Trail making A test, Mdn [IQR]		81 [70]	
Stroop effect, M [SD]		17.5 [9.9]	
VOSP Silhouettes, M [SD]		14.3 [5.7]	
Short Delay Cued Recall ^f , M [SD]		3.7 [2.3]	

M, mean; SD, standard deviation; Mdn, median; IQR, interquartile range; MMSE, Mini-Mental State Examination; BP, blood pressure; CIRS, Cumulative illness rating scale; MADRS, Montgomery Asberg Depression Rating Scale; VOSP, Visual Object and Space Perception Battery. ^aUnivariate statistical analysis by student *T*-test. ^bUnivariate statistical analysis by Pearson Chi-Square test. ^cUnivariate statistical analysis by Mann-Whitney U test. ^dCardiac disease = any of the following: known coronary heart disease, heart failure or atrial fibrillation. ^eCerebrovascular disease = previous stroke or transient ischemic attacks. ^fFrom the California Verbal Learning Test, Second Edition, part A. *Statistically significant difference <0.05, or **highly significant, <0.001.

$p=0.004$), 5-HT_{2C}R-abs ($r=0.25$, $p=0.0005$), 5-HT₇R-abs ($r=0.21$, $p=0.003$), Stab1-abs ($r=0.23$, $p=0.001$), C5aR₁-abs ($r=0.21$, $p=0.004$), and NGF-abs ($r=0.19$, $p=0.01$) less so. The sensitivity and specificity of VEGFR1-abs to distinguish AD from controls was 0.58 and 0.52, respectively.

Immune-assays for determining antibody levels: Coefficients of variation

Fourteen out of thirty-three antibodies measured had coefficients of variation <10% (good), fifteen below 20% (acceptable) and four were above 20% (poor). Of the antibodies increased in AD, 5-HT₇R-abs had a coefficient of variation >20%. A full list of coefficients of variation can be found in Supplementary Table 2.

Confounding analysis of the difference between AD and controls

We found no influence of storage time on antibody levels (data not shown), consistent with previous

studies [32]. To identify if the group differences in vascular risk and disease confounded the relationship between AD and antibodies, we compared multivariate models with and without these confounders (Table 2).

None of the antibodies met the 10% change in estimate (OR) criterion, but NGF-abs were majorly confounded. Antibody levels were not different between neuropathologically confirmed and clinically diagnose AD (p -values [0.37–0.86], MWUT).

Correlations between antibodies: Within-group analysis of patients with AD

The antibodies in AD (controls not shown) correlated (Spearman) with each other with weak, moderate, and strong correlation coefficients and formed hierarchical clusters (Fig. 3).

Many strong correlations between antibody levels will result in lack of statistical independence of all observations. This is likely due to sequence and/or structural homology in the extracellular sequences of the antigens targeted.

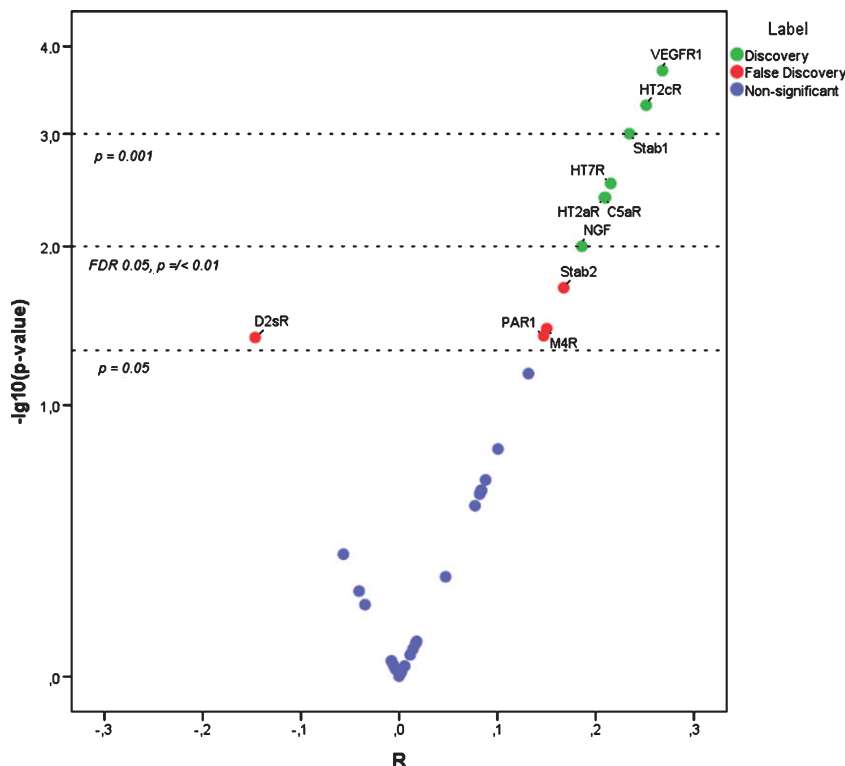


Fig. 1. Antibodies in Alzheimer's disease and healthy controls. *Non-parametric smile plot. The negative log of the p -value is on the y-axis (1 is $p = 0.1$, 2 is $p = 0.01$, 3 is $p = 0.001$) and R from Mann-Whitney U test (calculated as the test z -score/ \sqrt{N}). Positive R indicates association with AD. p -values of 0.05 and 0.001 and p -value cut-off adjusted for multiple testing with the FDR 0.05 are indicated.

Table 2
Multivariate analysis of Alzheimer's disease versus healthy controls

Antibodies to	Crude model ^a		Adjusted model ^b		
	OR	p	OR	Δ OR	p
VEGFR1	1.73	0.001	1.70	-0.02	0.004
5-HT _{2C} R	1.75	0.001	1.81	0.03	0.002
Stab1	1.65	0.002	1.54	-0.07	0.016
5-HT ₇ R	1.60	0.003	1.48	-0.07	0.023
C5aR ₁	1.51	0.008	1.57	0.04	0.013
5-HT _{2A} R	1.57	0.004	1.51	-0.04	0.024
NGF	1.40	0.025	1.37	-0.03	0.076*

OR, odds ratio; p , p -value; Δ OR = ((Crude OR - Adjusted OR)/Crude OR). Δ OR $> \pm 0.1$ is minor confounding. ^aOutcome: AD versus controls, covariates: age, gender and antibody as listed (logistic regression). ^bOutcome: AD versus controls, covariates: age, gender, cardiac disease, diabetes, stroke, hypertension, BP > 140 , current smoking and antibody as listed (logistic regressions). *Major confounding (change to non-significant p -value).

Antibodies, cognition, and mood: Within-group analysis of patients with AD

Age, gender, CIRS, and education (dichotomized > 10 years) were included as covariates in all regressions. All cognitive test results were transformed so that a higher score is worse (Fig. 3).

Psychomotor speed (TMT-A) was most strongly associated with D₁R-abs (β 0.49, $p < 0.001$), depression (MADRS) with ETAR-abs (β 0.31, $p < 0.001$), and visuospatial function (VOSP) with 5-HT_{1A}R-abs (β 0.27, $p = 0.004$). There were no discoveries with MMSE, memory (CVLTA-SDCR), or executive function (Stroop-E).

We did not identify major confounding of these associations. Some confounders were themselves associated the outcomes. Current smoking (β 0.43, $p = 0.03$) and CBVD (β 0.68, $p = 0.004$) were associated with TMT-A and low BMI with VOSP (β -0.35, $p = 0.03$). The full confounding analysis can be found in Supplementary Table 3.

DISCUSSION

AD was associated with a significant increase in the levels of six antibodies to receptors from the vasculature (VEGFR1-abs), immune (Stab1-abs, and C5aR₁-abs), and serotonergic systems (5-HT_{2A}R-abs, 5-HT_{2C}R-abs and 5-HT₇R-abs), after adjustment for multiple testing and differences in vascular

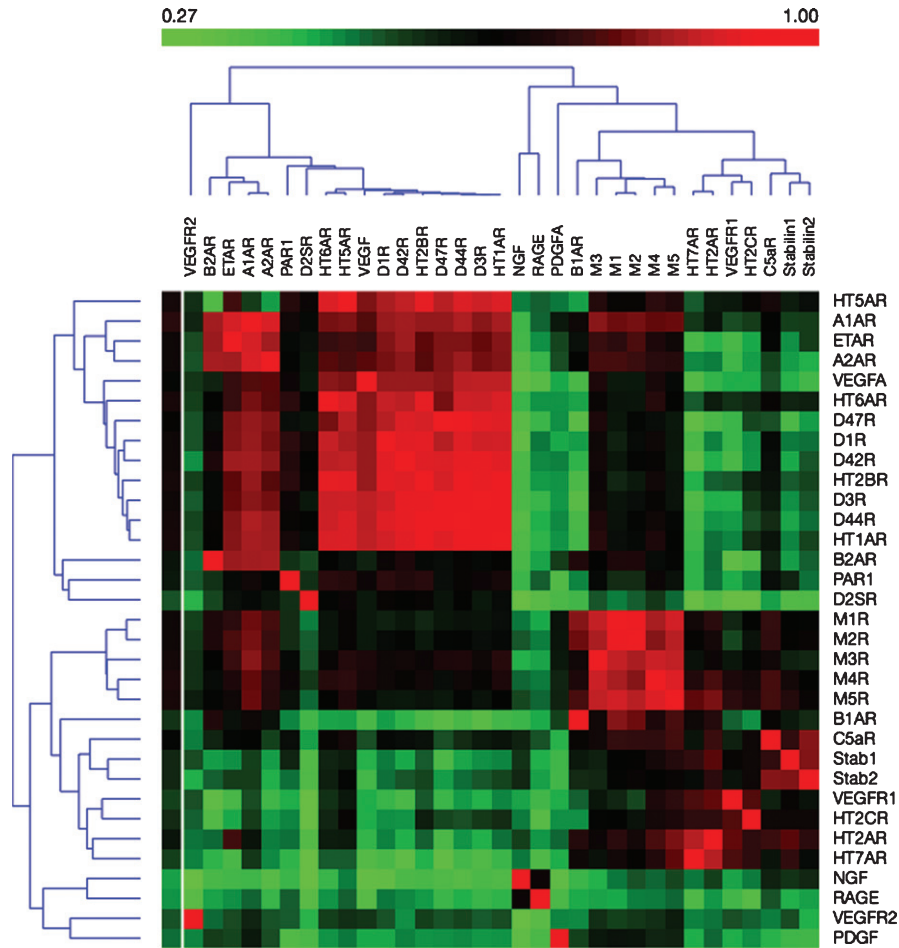


Fig. 2. Antibodies are correlated and form hierarchical clusters (heatmap). *Antibodies in hierarchical clusters by their spearman correlation coefficient. Weak correlations are displayed in green, moderate in black and strong in red.

co-morbidities. The difference in NGF-abs was confounded by group differences in vascular co-morbidities. Antibodies to VEGFR1 had the most significant association with AD. VEGFR1 is mainly expressed by the cerebral endothelium [33], but antibodies to other endothelial receptors were not increased. The main function of the innate immune system receptors C5aR₁ and Stab1 are initiation of inflammation and scavenging, respectively [34, 35]. The serotonergic receptors that were targeted by antibodies have their highest level of expressions in the brain [36].

The physiological mechanisms that lead to the formation of auto-IgG Nabs, including brain antigens, are not firmly established. The level of Nabs can rise in response to apoptosis of cells and oxidation of proteins [37]. There is increased migration of immune cells to the brain in AD, including T-cells

[38], monocytes [39], and neutrophils [40]. Expression of major histocompatibility protein 2, a key molecule for antigen-presentation, is increased in AD microglia [41]. This could generate an environment that favors autoantibody formation toward brain antigens. In line with this, antibodies to serotonergic receptors expressed mainly in the brain, were increased. Cholinergic neurons undergo extensive apoptosis in AD [42], but antibodies to muscarinic cholinergic receptors were not increased. Thus, we did not find convincing evidence to support AD-related cell-injury in the brain as a driver of antibody generation. More commonly, antibodies to brain antigens are initiated toward extra-cerebral antigens that are also expressed in the brain, or have shared epitopes with brain-antigens [43]. Apoptosis and oxidative stress also occur in the endothelial lining of brain [44, 45] and oxidative stress markers are

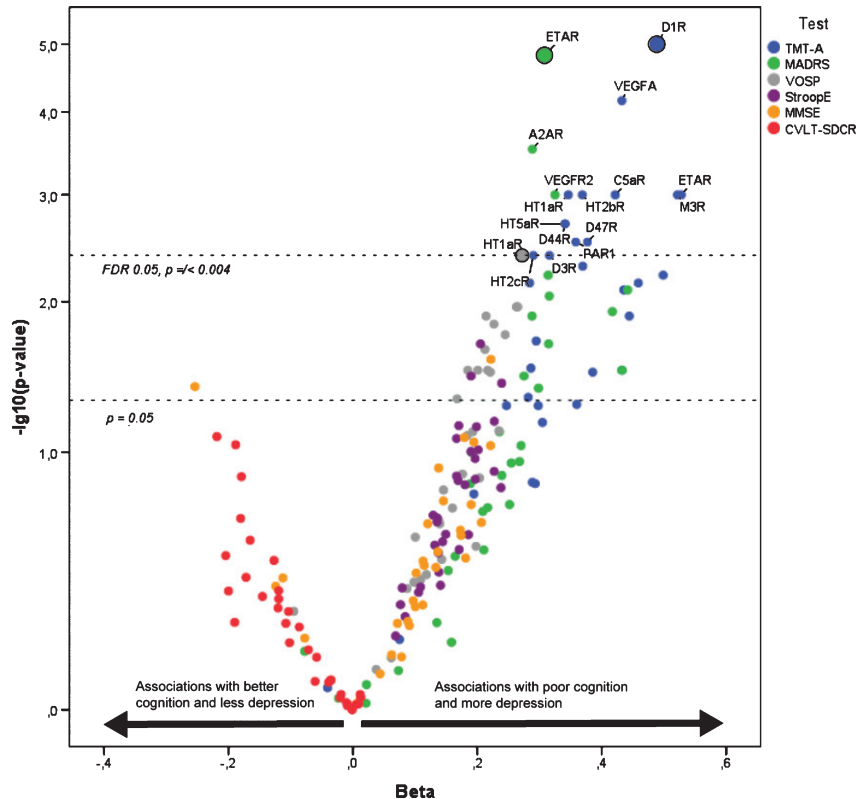


Fig. 3. Antibodies, cognitive testing, and depression. *Smile plot of 196 robust regressions by MM estimation where each antibody is analyzed with covariates and the cognitive test (color coded) as the outcome. $-\log(p\text{-value})$ on the Y axis generates a scale were (1 is $p=0.1$), (2 is $p=0.01$), (3 is $p=0.001$), and (4 is $p=0.0001$). Beta indicates standardized regression coefficients. Positive beta values indicate that antibodies are associated with poorer cognitive function and higher depression scores. The strongest associations with a particular cognitive test have larger dots. $p=0.05$ and the FDR cutoff at $FDR=0.05$ are marked by lines. All analyses are with age, gender, education, and CIRS in the equation. MMSE, Mini-Mental State Examination; CVLT-SDRC, CVLTA Short Delayed Cued Recall; Stroop E, Stroop effect; MADRS, Montgomery-Aasberg Depression Rating Scale; TMT-A, Trail Making A test. Three cases were missing for TMT-A, 1 for VOSP, and 3 for Stroop effect.

generally increased in AD [46]. The antigens targeted are expressed at low to moderate levels in brain microvascular cells and immune-cells [36]. It is perhaps more likely that antibody formation is initiated due to AD-related cellular damage to cells outside the brain.

As illustrated in Fig. 2, we observed high correlations between the antibodies. Immunization studies have found the immune-assays applied here to be specific when animals are immunized to a linear epitope [47]. Nabs do not have the same high affinity binding to antigens as antibodies raised by immunization and are typically polyreactive. We propose that the observed correlations represent polyreactivity with linear and/or conformational epitopes. In support of this, the highest correlations between antibodies were between different dopaminergic antibodies ($r>0.9$), which do have high structural

similarity [48]. This suggests that the antibodies measured are Nabs, a physiological part of the innate immune system.

Poor cognitive performance and higher depression scores were associated with levels of antibodies that were not significantly increased in AD compared to controls. The strongest associations were between slower psychomotor speed (TMT-A) and D₁R-abs (moderate effect), more depression (MADRS) and ETAR-abs (moderate effect) and finally worse visuospatial function (VOSP) and 5-HT_{1A}R-abs (small effect). These three associations were not confounded by the ApoE4 genotype, leukocyte levels, BMI, or vascular risk factors or disease. AD patients that were current smokers or had CBVD also had slower psychomotor speed, while a high BMI was associated with better visuospatial function (Supplementary Table 3).

D₁R, 5-HT_{1A}R and ETAR have moderate to high expression in the brain [49–51]. Brain-reactive antibodies are found in healthy individuals and the antibodies observed against D₁R and 5-HT_{1A}R could represent brain-reactive antibodies at physiological levels [52]. Decreased D₁-receptor expression is associated with cognitive slowing [53] and 5-HT_{1A}-receptors are involved in spatial learning and memory [54]. ETAR does not have an established role in depression, but ETAR antagonists had a beneficial effect on depression in an animal model by influencing cytokine secretion [55].

The association of antibodies at physiological levels with poor cognitive function and depression may indicate that AD-patients could have an underlying frailty to antibodies directed at certain receptors, though we do not know if similar associations were present among the controls. It has been suggested that healthy elderly individuals have NMDAR-abs with pathogenic potential and that this pathogenic potential can become realized when there is BBB breakdown [9]. Alternatively, our findings could be related to an increase in Nabs due to a pro-inflammatory state. BBB damage could lead to more brain-immune interactions in AD than occurs under physiological conditions [56]. Levels of CRP are linked to both psychomotor speed and depression [57, 58]. In AD, neuropsychiatric symptoms are linked to levels of cytokines [59], despite no increase in levels of most cytokines in AD [60]. Our findings need to be investigated in future studies for validation and exploration of any potential mechanisms underlying the associations between serum-antibodies, cognitive function, and mood in AD. Our study lacked neuropsychological tests for the healthy controls so we could not estimate any associations between Nabs and cognitive function and depression in healthy individuals. While coefficients of variations were acceptable for the antibodies associated with AD and cognitive tests, 5-HT₇R was an exception with an inter-assay variability >20% and this finding should thus be interpreted with caution. Further studies are necessary to identify antibody targets and the functional consequences of antibody binding. To adequately address any effect modification of underlying BBB impairment, the degree of BBB compromise and intra-thecal antibody levels needs to be measured. Our study had adequate power to test all the primary hypotheses and conduct confounding analyses, but was underpowered to test for effect modification. Strengths in our study include the standardized clinical and biomarker assessments of the AD patients and autopsy diagnosis

in a subset. The overall diagnostic accuracy is thus likely to be high.

In summary, AD was associated with increased levels of Nabs targeting receptors of the innate immune system (Stabilin-1 and C5aR₁), of brain microvascular cells (VEGFR1) and serotonergic receptors (5-HT_{2A}R, 5-HT_{2C}R, and 5-HT₇R), though effect sizes were small. Nabs at physiological levels were associated with cognition and mood in the AD group: ETAR-abs with depression (MADRS, moderate effect), D₁R-abs with psychomotor speed (TMT-A, moderate effect), and 5-HT_{1A}R-abs with visuospatial function (VOSP, small effect).

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SUPPLEMENTARY MATERIAL

The supplementary material is available in the electronic version of this article: <http://dx.doi.org/10.3233/JAD-170245>.

REFERENCES

- [1] Ritchie K, Lovestone S. The dementias. *Lancet* **360**, 1759-1766.
- [2] Ittner LM, Götz J (2011) Amyloid- β and tau — a toxic pair de deux in Alzheimer's disease. *Nat Rev Neurosci* **12**, 67-72.
- [3] Mattson MP (2000) Apoptosis in neurodegenerative disorders. *Nat Rev Mol Cell Biol* **1**, 120-130.
- [4] Selkoe DJ (2002) Alzheimer's disease is a synaptic failure. *Science* **298**, 789-791.
- [5] Querfurth HW, LaFerla FM (2010) Alzheimer's disease. *N Engl J Med* **362**, 329-344.
- [6] Lutz HU (2007) Homeostatic roles of naturally occurring antibodies: An overview. *J Autoimmun* **29**, 287-294.

- [7] Nagele EP, Han M, Acharya NK, DeMarshall C, Kosciuk MC, Nagele RG (2013) Natural IgG autoantibodies are abundant and ubiquitous in human sera, and their number is influenced by age, gender, and disease. *PLoS One* **8**, e60726.
- [8] Silverman GJ, Gronwall C, Vas J, Chen Y (2009) Natural autoantibodies to apoptotic cell membranes regulate fundamental innate immune functions and suppress inflammation. *Discov Med* **8**, 151-156.
- [9] Castillo-Gomez E, Oliveira B, Tapken D, Bertrand S, Klein-Schmidt C, Pan H, Zafeiriou P, Steiner J, Jurek B, Trippe R, Pruss H, Zimmermann WH, Bertrand D, Ehrenreich H, Hollmann M (2016) All naturally occurring autoantibodies against the NMDA receptor subunit NR1 have pathogenic potential irrespective of epitope and immunoglobulin class. *Mol Psychiatry*, doi: 10.1038/mp.2016.1125
- [10] Diamond B, Honig G, Mader S, Brimberg L, Volpe BT (2013) Brain-reactive antibodies and disease. *Annu Rev Immunol* **31**, 345-385.
- [11] Bowman GL, Quinn JF (2008) Alzheimer's disease and the blood-brain barrier: past, present and future. *Aging Health* **4**, 47-55.
- [12] Wu J, Li L (2016) Autoantibodies in Alzheimer's disease: Potential biomarkers, pathogenic roles, and therapeutic implications. *J Biomed Res* **30**, 361-372.
- [13] Giil LM, Kristoffersen EK, Vedeler CA, Aarsland D, Nordrehaug JE, Winblad B, Cedazo-Minguez A, Lund A, Reksten TR (2015) Autoantibodies toward the angiotensin 2 type 1 receptor: A novel autoantibody in Alzheimer's disease. *J Alzheimers Dis* **47**, 523-529.
- [14] Wu J, Li L (2016) Autoantibodies in Alzheimer's disease: Potential biomarkers, pathogenic roles, and therapeutic implications. *J Biomed Res* **30**, 361-372.
- [15] Karczewski P, Hempel P, Kunze R, Bimmler M (2012) Agonistic autoantibodies to the alpha(1)-adrenergic receptor and the beta(2)-adrenergic receptor in Alzheimer's and vascular dementia. *Scand J Immunol* **75**, 524-530.
- [16] Busse S, Brix B, Kunschmann R, Bogerts B, Stoecker W, Busse M (2014) N-methyl-D-aspartate glutamate receptor (NMDA-R) antibodies in mild cognitive impairment and dementias. *Neurosci Res* **85**, 58-64.
- [17] Aarsland D, Rongve A, Nore SP, Skogseth R, Skulstad S, Ehrt U, Hoprekstad D, Ballard C (2008) Frequency and case identification of dementia with Lewy bodies using the revised consensus criteria. *Dement Geriatr Cogn Disord* **26**, 445-452.
- [18] Jack CR Jr, Albert MS, Knopman DS, McKhann GM, Sperling RA, Carrillo MC, Thies B, Phelps CH (2011) Introduction to the recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* **7**, 257-262.
- [19] Oppedal K, Aarsland D, Firbank MJ, Sonnesyn H, Tysnes OB, O'Brien JT, Beyer MK (2012) White matter hyperintensities in mild Lewy body dementia. *Dement Geriatr Cogn Dis Extra* **2**, 481-495.
- [20] Huntley AL, Johnson R, Purdy S, Valderas JM, Salisbury C (2012) Measures of multimorbidity and morbidity burden for use in primary care and community settings: A systematic review and guide. *Ann Fam Med* **10**, 134-141.
- [21] Berge G, Sando SB, Rongve A, Aarsland D, White LR (2014) Apolipoprotein E epsilon2 genotype delays onset of dementia with Lewy bodies in a Norwegian cohort. *J Neurol Neurosurg Psychiatry* **85**, 1227-1231.
- [22] Loebel M, Grabowski P, Heidecke H, Bauer S, Hanitsch LG, Wittke K, Meisel C, Reinke P, Volk HD, Fluge O, Mella O, Scheibenbogen C (2016) Antibodies to beta adrenergic and muscarinic cholinergic receptors in patients with chronic fatigue syndrome. *Brain Behav Immun* **52**, 32-39.
- [23] Lee H, Whitfield PL, Mackay CR (2008) Receptors for complement C5a. The importance of C5aR and the enigmatic role of C5L2. *Immunol Cell Biol* **86**, 153-160.
- [24] Tuszynski MH, Yang JH, Barba D, U HS, Bakay RA, Pay MM, Masliah E, Conner JM, Kobalka P, Roy S, Nagahara AH (2015) Nerve growth factor gene therapy: Activation of neuronal responses in Alzheimer disease. *JAMA Neurol* **72**, 1139-1147.
- [25] Provias J, Jaynes B (2014) The role of the blood-brain barrier in the pathogenesis of senile plaques in Alzheimer's disease. *Int J Alzheimers Dis* **2014**, 191863.
- [26] Sanchez-Cubillo I, Perianez JA, Adrover-Roig D, Rodriguez-Sanchez JM, Rios-Lago M, Tirapu J, Barcelo F (2009) Construct validity of the Trail Making Test: Role of task-switching, working memory, inhibition/interference control, and visuospatial abilities. *J Int Neuropsychol Soc* **15**, 438-450.
- [27] Muller-Thomsen T, Arlt S, Mann U, Mass R, Ganzer S (2005) Detecting depression in Alzheimer's disease: Evaluation of four different scales. *Arch Clin Neuropsychol* **20**, 271-276.
- [28] Brookes ST, Whitely E, Egger M, Smith GD, Mulheran PA, Peters TJ (2004) Subgroup analyses in randomized trials: Risks of subgroup-specific analyses; power and sample size for the interaction test. *J Clin Epidemiol* **57**, 229-236.
- [29] Faul F, Erdfelder E, Lang AG, Buchner A (2007) G*Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods* **39**, 175-191.
- [30] Yohai VJ (1987) High breakdown-point and high efficiency robust estimates for regression. *Ann Stat* **15**, 642-656.
- [31] Demsar J, Curk T, Erjavec A, Gorup C, Hocevar T, Milutinovic M, Mozina M, Polajnar M, Toplak M, Staric A, Stajdohar M, Umek L, Zagar L, Zbonart J, Zitnik M, Zupan B (2013) Orange: Data mining toolbox in python. *J Mach Learn Res* **14**, 2349-2353.
- [32] Petrakis NL (1985) Biologic banking in cohort studies, with special reference to blood. *Natl Cancer Inst Monogr* **67**, 193-198.
- [33] Witmer AN, Dai J, Weich HA, Vrensen GF, Schlingemann RO (2002) Expression of vascular endothelial growth factor receptors 1, 2, and 3 in quiescent endothelia. *J Histochem Cytochem* **50**, 767-777.
- [34] Ward PA (2009) Functions of C5a receptors. *J Mol Med (Berl)* **87**, 375-378.
- [35] Kzhyshkowska J (2010) Multifunctional receptor stabilin-1 in homeostasis and disease. *ScientificWorldJournal* **10**, 2039-2053.
- [36] Uhlen M, Fagerberg L, Hallstrom BM, Lindskog C, Oksvold P, Mardinoglu A, Sivertsson A, Kampf C, Sjostedt E, Asplund A, Olsson I, Edlund K, Lundberg E, Navani S, Szgyarto CA, Odeberg J, Djureinovic D, Takanen JO, Hober S, Alm T, Edqvist PH, Berling H, Tegel H, Mulder J, Rockberg J, Nilsson P, Schwenk JM, Hamsten M, von Feilitzen K, Forsberg M, Persson L, Johansson F, Zwaalen M, von Heijne G, Nielsen J, Ponten F (2015) Proteomics. Tissue-based map of the human proteome. *Science* **347**, 1260419.

- [37] Chang MK, Binder CJ, Miller YI, Subbanagounder G, Silverman GJ, Berliner JA, Witztum JL (2004) Apoptotic cells with oxidation-specific epitopes are immunogenic and proinflammatory. *J Exp Med* **200**, 1359-1370.
- [38] Town T, Tan J, Flavell RA, Mullan M (2005) T-cells in Alzheimer's disease. *Neuromolecular Med* **7**, 255-264.
- [39] Saresella M, Marventano I, Calabrese E, Piancone F, Rainone V, Gatti A, Alberoni M, Nemni R, Clerici M (2014) A complex proinflammatory role for peripheral monocytes in Alzheimer's disease. *J Alzheimers Dis* **38**, 403-413.
- [40] Zenaro E, Pietronigro E, Della Bianca V, Piacentino G, Marongiu L, Budui S, Turano E, Rossi B, Angiari S, Dusi S, Montesor A, Carlucci T, Nani S, Tosadori G, Calciano L, Catalucci D, Berton G, Bonetti B, Constantin G (2015) Neutrophils promote Alzheimer's disease-like pathology and cognitive decline via LFA-1 integrin. *Nat Med* **21**, 880-886.
- [41] Hofsfield LA, Humpel C (2015) Migration of blood cells to beta-amyloid plaques in Alzheimer's disease. *Exp Gerontol* **65**, 8-15.
- [42] Arendt T, Bruckner MK, Morawski M, Jager C, Gertz HJ (2015) Early neurone loss in Alzheimer's disease: Cortical or subcortical? *Acta Neuropathol Commun* **3**, 10.
- [43] Korn T, Kallies A (2017) T cell responses in the central nervous system. *Nat Rev Immunol* **17**, 179-194.
- [44] Massaad CA (2011) Neuronal and vascular oxidative stress in Alzheimer's disease. *Curr Neuropharmacol* **9**, 662-673.
- [45] Religa P, Cao R, Religa D, Xue Y, Bogdanovic N, Westaway D, Marti HH, Winblad B, Cao Y (2013) VEGF significantly restores impaired memory behavior in Alzheimer's mice by improvement of vascular survival. *Sci Rep* **3**, 2053.
- [46] Schrag M, Mueller C, Zabel M, Crofton A, Kirsch WM, Ghribi O, Squitti R, Perry G (2013) Oxidative stress in blood in Alzheimer's disease and mild cognitive impairment: A meta-analysis. *Neurobiol Dis* **59**, 100-110.
- [47] Wenzel K, Rajakumar A, Haase H, Geusens N, Hubner N, Schulz H, Brewer J, Roberts L, Hubel CA, Herse F, Hering L, Qadri F, Lindschau C, Wallukat G, Pijnenborg R, Heidecke H, Riemekasten G, Luft FC, Muller DN, Lamarca B, Dechend R (2011) Angiotensin II type 1 receptor antibodies and increased angiotensin II sensitivity in pregnant rats. *Hypertension* **58**, 77-84.
- [48] Platania CB, Salomone S, Leggio GM, Drago F, Bucolo C (2012) Homology modeling of dopamine D2 and D3 receptors: Molecular dynamics refinement and docking evaluation. *PLoS One* **7**, e44316.
- [49] Meador-Woodruff JH, Damask SP, Wang J, Haroutunian V, Davis KL, Watson SJ (1996) Dopamine receptor mRNA expression in human striatum and neocortex. *Neuropsychopharmacology* **15**, 17-29.
- [50] Naidoo V, Naidoo S, Mahabeer R, Raidoo DM (2004) Cellular distribution of the endothelin system in the human brain. *J Chem Neuroanat* **27**, 87-98.
- [51] Miguelez C, Morera-Herrerias T, Torrecilla M, Ruiz-Ortega JA, Ugedo L (2014) Interaction between the 5-HT system and the basal ganglia: Functional implication and therapeutic perspective in Parkinson's disease. *Front Neural Circuits* **8**, 21.
- [52] Levin EC, Acharya NK, Han M, Zavareh SB, Sedeyn JC, Venkataraman V, Nagele RG (2010) Brain-reactive autoantibodies are nearly ubiquitous in human sera and may be linked to pathology in the context of blood-brain barrier breakdown. *Brain Res* **1345**, 221-232.
- [53] Rieckmann A, Karlsson S, Karlsson P, Brehmer Y, Fischer H, Farde L, Nyberg L, Backman L (2011) Dopamine D1 receptor associations within and between dopaminergic pathways in younger and elderly adults: Links to cognitive performance. *Cereb Cortex* **21**, 2023-2032.
- [54] Gliemann-Johnston Y, Saling MM, Reutens DC, Stout JC (2015) Hippocampal 5-HT1A receptor and spatial learning and memory. *Front Pharmacol* **6**, 289.
- [55] Pinho-Ribeiro FA, Borghi SM, Staurengo-Ferrari L, Filgueiras GB, Estanislau C, Verri WA Jr (2014) Bosentan, a mixed endothelin receptor antagonist, induces antidepressant-like activity in mice. *Neurosci Lett* **560**, 57-61.
- [56] Zenaro E, Piacentino G, Constantin G (2016) The blood-brain barrier in Alzheimer's disease. *Neurobiol Dis*, doi: 10.1016/j.nbd.2016.07.007
- [57] Wium-Andersen MK, Orsted DD, Nielsen SF, Nordestgaard BG (2013) Elevated C-reactive protein levels, psychological distress, and depression in 73, 131 individuals. *JAMA Psychiatry* **70**, 176-184.
- [58] Hoth KF, Haley AP, Gunstad J, Paul RH, Poppas A, Jefferson AL, Tate DF, Ono M, Jerskey BA, Cohen RA (2008) Elevated C-reactive protein is related to cognitive decline in older adults with cardiovascular disease. *J Am Geriatr Soc* **56**, 1898-1903.
- [59] Holmes C, Cunningham C, Zotova E, Culliford D, Perry VH (2011) Proinflammatory cytokines, sickness behavior, and Alzheimer disease. *Neurology* **77**, 212-218.
- [60] Brosseron F, Krauthausen M, Kummer M, Heneka MT (2014) Body fluid cytokine levels in mild cognitive impairment and Alzheimer's disease: A comparative overview. *Mol Neurobiol* **50**, 534-544.