#### Introduction

The CellTrend IgA(rabbit)-ELISA is designed for the quantitative determination of rabbit IgA in complex samples (e.g. serum or other biological samples).

## **Principle of the Assay**

The determination of rabbit IgA is carried out as direct sandwich ELISA. An antibody specific for rabbit IgA has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any IgA present is bound. After washing away any unbound substances, an enzyme-linked antibody is added. Following a wash, a substrate solution is added to the wells and color develops in proportion to the amount of antibody conjugate. The absorption at 450 nm is proportional to the IgA concentration.

#### **Precautions**

Store the kit at 2-8 °C.

For research use only. Not for use in diagnostic procedures.

For in vitro use only.

Do not use the reagents beyond the expiration date marked on box label.

Please read the instructions carefully before using the kit.

Do not mix reagents from different lots.

Some components of this kit contain Thimerosal, a mercury containing compound. The stop solution contains 0.5 M sulphuric acid. Follow routine precautions for handling hazardous chemicals.

# Other supplies required

Deionized or distilled water Graduated cylinder Micropipettes, multipipette Microplate reader

## **Preparation of reagents and samples**

- Bring all reagents to room temperature before use. If crystals have formed, mix gently until the crystals have completely dissolved.
- The <u>microplate strips</u> are ready to use. Remove excess strips (breakable) from the frame, reseal in the bag with the desiccant and store at 2-8 °C
- Dilute the <u>wash buffer</u> with deionized or distilled water **1:10** (e. g. 40 ml + 360 ml water). The diluted solution is stable for 30 days at 2-8 °C.
- Use the <u>Standard concentrate</u> to produce a 1:2-dilution series with diluent (e. g. 250  $\mu$ l + 250  $\mu$ l diluent):

standard	preparation	conc. ng/ml
S 7	standard conc. undiluted	1000.000
S 6	S 7 1:2 diluted	500.000
S 5	S 6 1:2 diluted	250.000
S 4	S 5 1:2 diluted	125.000
S 3	S 4 1:2 diluted	62.500
S 2	S 3 1:2 diluted	31.250
S 1	S 2 1:2 diluted	15.625

- Dilute the <u>samples</u> with diluent. If samples generate values outside the standard curve, the dilution factor may be varied. To exclude matrix effects the dilution factor should be at least 1:50.

#### Assay procedure

It is recommended that all samples and standards be assayed in duplicate.

- Prepare all reagents, standard curve and samples as directed in the previous section.
- Pipette 100 µl of samples, standards, positive control or diluent (as negative control) into the wells.
- 3. Seal wells with adhesive strip and incubate for 1 hour at room temperature with shaking \*.
- Remove fluid from wells and wash three times with 300 μl wash buffer. After the last wash, invert the plate and tap on a clean paper towel.
- 5. Add 100 μl of HRP conjugate to each well.
- 6. Seal wells with adhesive strip and incubate for 1 hour at room temperature with shaking \*.
- 7. Repeat the wash as in step 4.
- 8. Dispense 100 µl of TMB substrate solution into each well.
- 9. Incubate for 10 minutes at room temperature in the dark.
- 10. Add 100 µl of stop solution to each well.
- 11. Determine the absorbance within 30 minutes at 450 nm. A reference wavelength of 620 nm/690 nm is recommended.

#### Calculation of results

Create a standard curve using computer software capable of generating a curve fit (4 parameter fit; x-axis: log, IgA concentration; y-axis: linear, absorbance). As an alternative, draw a standard curve on semi-log paper (x-axis: log, IgA concentration; y-axis: linear, absorbance). The IgA concentrations can be calculated from the standard curve. The calculated concentrations must be multiplied by the sample dilution factor.

If the absorbance of some samples is outside the standard curve a subsequent determination with changed dilutions will provide a proper result.

<sup>\*:</sup> If a MTP shaker is not available, it is possible to incubate for 2 hours without shaking.

## **Materials provided:**

Number of determinations/Catalog No.	1x96 determ. 56100
Microplate strips, antibody coated	12 x 8
Wash buffer, 10fold conc. ◆	50 ml
Diluent, ready to use ◆	100 ml
Standard concentrate, 1000 ng/ml ◆	2 ml
Anti-IgA(rabbit)-Ab., HRP conjugate, ready to use	12 ml
TMB substrate, ready to use	12 ml
Stop solution, ready to use (0.5 M sulphuric acid)	12 ml

<sup>♦:</sup> contains Thimerosal

## Assay procedure summary:

#### A. Preparation

- 1. Bring all reagents to room temperature
- 2. Dilute wash buffer 1:10
- 3. Prepare the standard curve from a 1:2-dilution series of standard concentrate with diluent
- 4. Dilute samples with diluent

## B. Performance

- 1. Pipette 100 µl of samples, standards, controls into the wells
- 2. Incubate for 1 hour at room temperature with shaking
- 3. Wash three times with 300 µl of wash buffer
- 4. Add 100 μl of HRP conjugate to each well
- 5. Incubate for 1 hour at room temperature with shaking
- 6. Wash three times with 300 µl of wash buffer
- 7. Dispense 100 µl of TMB substrate solution
- 8. Incubate for 10 minutes at room temperature in the dark
- 9. Add 100 µl of stop solution
- 10. Measure absorption at 450 nm

# **Related products:**

ELISA for quantitative determination of IgG (Rabbit) (Catalog number: 56100) ELISA for quantitative determination of IgM (Rabbit) (Catalog number: 56200)

Contract analysis: determination of IgG/IgA/IgM in rabbit samples (please inquire)

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#### INSTRUCTIONS FOR USE

# ELISA for Quantitative Determination of IgA (Rabbit)

Catalog No.: 68100



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